Multi-functional Roles of Granulocyte Macrophage Colony-Stimulating Factor in Atherosclerosis

Yasuyuki Sasaguri*, and Akihide Tanimoto**

Granulocyte macrophage colony-stimulating factor (GM-CSF) is a growth factor for hematopoietic progenitor cells, and is one of major pro-inflammatory cytokines. In the atherosclerotic lesion, the cells constituting the arterial intima such as endothelial and smooth muscle cells, and monocytes/macrophages can produce GM-CSF in response to other cytokines, modified low-density lipoprotein and cell-cell interaction. GM-CSF exhibits multi-functional effects on the intimal cells and closely related to the pathogenesis of atherosclerosis. In particular, the monocytes/macrophages are present in all stages of atherosclerosis and play a central role in atherogenesis. Monocytic differentiation, macrophages proliferation, expression of adhesion molecules, matrix-degrading enzymes and cytokines are regulated by GM-CSF. Among these, monocytic migration and matrix metalloproteinase expression would be one of critical functions for monocytes/macrophages. GM-CSF is also related to angiogenesis and vasculogenesis by enhancing endothelial cell mobilization from progenitor cells and monocyte adhesion(J. Jpn. Coll. Angiol., 2003, 43: 715-723)

Key words: GM-CSF, Atherosclerosis, Monocytes/macrophages, Migration, Matrix metalloproteinase (MMP)

Introduction

Inflammatory events are involved in the pathogenesis of atherosclerosis1~2), and T-lymphocytes, monocytes/macrophages, and monocyte-derived dendritic cells are abundant in the atherosclerotic lesion^{3~5}). These inflammatory cells are also functional to regulate Th-1 and Th-2 balance in the lesion environment of atherosclerosis^{6~7}). Especially, monocytes/macrophages are present in all stages of atherosclerosis, playing an important role in atherogenesis; and they have multiple functions, which include migration and secretion of growth factors, cytokines, and matrix-degrading enzymes, and the uptake of modified lipoproteins^{8~9}). During the initiation of atherosclerosis or inflammation, circulating monocytes adhere to endothelial cells via specific cell-adhesion molecules, and subsequently migrate into the subendothelial space¹⁰). These processes take place in the micro-environment formed by endothelium, intimal smooth muscle cells, migrating lymphocytes and macrophages, and extracellular matrixs, as well as being regulated by many biologically active substances secreted from the constituent¹).

In this review we describe the multi-functional roles of granulocyte macrophage colony-stimulating factor (GM-CSF) in relation to atherosclerosis. Especially, we focus on GM-CSF effects for monocyte/macrophage functions, including migration and matrix metalloproteinase (MMP) expression. Furthermore, the importance of GM-CSF in monocyte/ macrophage functions is more emphasized than that of platelet-derived growth factor (PDGF), which is a predominant factor for the functions of smooth muscle cells (SMC).

1. Multi-functional Role of GM-CSF

GM-CSF is a growth factor required for the survival, growth, and differentiation of hematopoietic proenitor cells¹¹, and is produced by inflammation-related cells including activated T-cells, monocytes/macrophages, and endothelial cells¹². GM-CSF is involved in a cytokine/growth factor network present in the atherosclerotic lesion^{1, 13-14}. With regards to monocyte/macrophage functions, GM-CSF plays an essen-

^{*} Department of Pathology and Cell Biology, School of Medicine, University of Occupational and Environmental Health, Kitakyushu, Japan

^{**} Department of Pathology, Toranomon Hpospital and Okinaka Memorial Institute for Medical Research, Tokyo, Japan

Accepted August 6, 2003

tial role in monocyte differentiation¹²) and macrophage proliferation¹⁵⁻¹⁶) in relation to the pathogenesis of atherosclerosis. In addition, the expression of adhesion molecules, MMPs, plasminogen-activator inhibitor, urokinase-type plasminogen activator, and myeloperoxidase is regulated by GM-CSF in monocytes/macrophages^{14, 17-20}). These findings indicate that GM-CSF modulates the conditions of the lesion environment of atherosclerosis.

2. Rho activation and GM-CSF

The Rho family GTPases, including Rho (A, B and C), Rac, and Cdc42, have been found to mediate cell adhesion, migration, and invasion. Several lines of evidence have suggested that the Rho is directly linked to the migratory or invasive phenotypes of epithelium-derived tumor cells²¹). In inflammation, the first step of leukocyte/monocyte transmigration is an interaction with endothelial cells; and the activated endothelial cells express adhesion molecules including E-selectin, intercellular adhesion molecule-1 (ICAM-1), and vascular cell adhesion molecule-1 (VCAM-1)²²). The clustering of these cell-surface molecules induced by monocyte adhesion is regulated by the Rho signaling pathway²³). RhoA is also activated after the stimulation with GM-CSF (unpublished data).

The signals for monocyte chemoattractant protein-1 (MCP-1)-dependent integrin activation via Rho are mediated by MCP-1 receptor CCR2, a member of the family of transmembrane-domain, G protein-coupled receptors²⁴). On the other hand, the growth factor receptors of receptor tyrosine kinase family, such as receptors for epidermal growth factor (EGF), hepatocyte growth factor (HGF), and PDGF-BB, activate Rho small G-protein via Ras activation²¹). Our results demonstrate that GM-CSF also stimulated the monocytes to activate RhoA and integrin clustering just like MCP-1 and platelet-derived growth factor-BB (PDGF-BB). At present, the molecular mechanism of the signaling pathway for GM-CSF-induced Rho activation is unknown. However, the GM-CSF-induced activation of a number of signaling molecules, including JAK2, Ras, Raf, ERK, and RSK²⁵ indicate a possible involvement of one of these signaling molecules as a mediator for Rho activation.

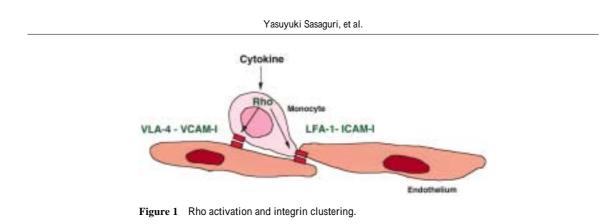
3. Integrin and GM-CSF

The recruitment of monocytes from the blood stream is one of the critical steps for atherosclerosis, and is regulated by various adhesion molecules expressed on both endothelial and monocytes¹). As shown in **Fig 1**, in monocytes/macrophages, very late antigen-4 (VLA-4) and lymphocyte function-associated antigen (LFA-1) are essential for facilitating lateral migration and diapedesis across the endothelial cells in the vascular wall, respectively^{26–27}). This monocyte-endothelial cell interaction is an initial event for atherogenesis. By stimulation with GM-CSF, both integrins are clustered on the cell surface (**Fig. 2**), indicating that GM-CSF, as well as PDGF-BB and monocyte MCP-1, is an important factor for the initial step of monocyte transendothelial migration.

4. Matrix metalloproteinase production and GM-CSF

A close linkage between MMP production and cell migratory activity of many types of cells may be implied; and the secretion of matrix-degrading enzymes, such as MMP, from monocytes is one of the key events to enhance the migratory action of the monocytes^{28 - 29}). GM-CSF has been reported to enhance MMP-12 expression in monocytes/macrophages, and the MMP-12 is localized in the atherosclerotic lesion^{17, 30}). Addition to MMP-12, MMP-1 and MMP-9 expression is also induced by GM-CSF in U937 monocytes (unpublished data) (**Fig. 3**). The expression of these MMPs in the atherosclerotic lesion is seen in commercially available antibodies (MMP-1, -7, and -9). For example, **Fig 4** shows the expression of MMP-1 and MMP-9 in monocyte-derived foam cells in the atherosclerosis. An overall review on MMP expression and atherogenesis is discussed in our previous review³¹).

GM-CSF induces MMP-9 expression in human peripheral blood monocytes³²), but the molecular mechanism for gene regulation by GM-CSF has not been clarified. The results of an earlier study using U937 monocytic cells indicated that the AP-1 motif in the MMP-12 promoter region was involved in gene regulation by GM-CSF, and that the AP-1 binding complex consisted of multiple fos/jun isoforms including junD, c-jun, fosB, c-fos, and fra-1¹⁷). The AP-1



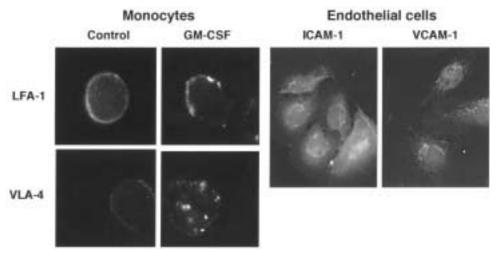


Figure 2 Immunofluorescence of integrin in monocytes (U937) and in human aortic endothelial cells. Left: After treatemtnt with GM-CSF, clustering of LFA-1 and VLA-4 (granular immunofluorescence) is taken place on the surface of monocytes.

Right: Upon stimulation with inflammatory stimuli (histamine), the expression of ICAM-1 and VCAM-1 is enhanced in the endothelial cells.

site is involved in many types of MMP gene expression^{33)} (Fig. 5). Recent report demonstrates an expression of extra-

cellular matrix metalloproteinase inducer (EMMPRIN) in human atheroma, and GM-CSF-induced macrophages show enhanced expression of the EMMPRIN as well as MMP-9³⁴). GM-CSF may play a central role monocytic MMP expression and its regulation in relation to atherogenesis.

PDGF, which has multilfunctional effects on SMC, does not enhance MMP-1, MMP-9 and MMP-12 expression in U937 monocytes (unpublished data).

5. Cytokine network and GM-CSF in atherosclerosis

In the atherosclerotic lesion environment, a cytokine/

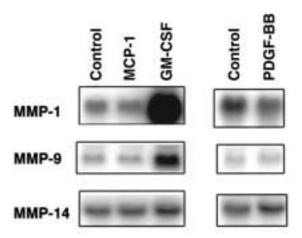


Figure 3 Northern bot analysis of MMPs mRNA. Unlike PDGF-BB, GM-CSF stimulates monocytes to up-regulated the transcrition of MMP-1 and MMP-9.

November, 25, 2003

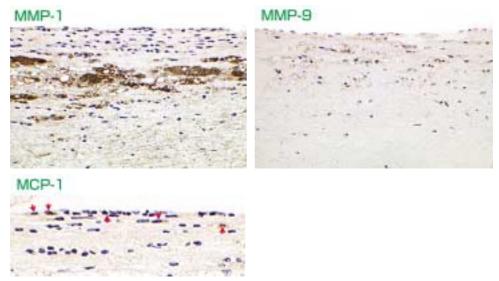
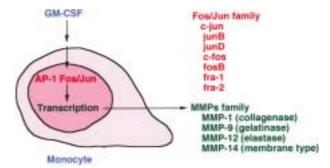


Figure 4 Immunohistochemical detection of MMP-1, MMP-9 and MCP-1 in human atherosclerotic lesions. MMP-1 and MMP-9 expresse in macrophages (form cells) in atherosclerotic thickened intima. MCP-1 is detected in endothelial cells and mononuclear cells beneath the endothelium (arrows).

growth factor network is constructed by various cytokines, growth factor, chemokines, and other biologically active substances^{35–36}). In this cytokine/ growth factor network, "cross talk" between various active substances determines the net effects of the cytokines. Among these cytokines or growth factors, we focus on the effects of GM-CSF in the monocytes, because some earlier reports demonstrated that GM-CSF stimulated peritoneal macrophages or a monocytic cell line to express MCP-1³⁷). Our data also shows that GM-CSF, but not PDGF, enhances MCP-

1 production at transcription, as well as mRNA and protein levels (unpublished data) (**Fig. 6**). GM-CSF may enhance monocytic functions via stimulation of MCP-1 production. GM-CSF has been reported only to prime monocytes for enhanced migration in response to MCP-1 through LFA-1 activation¹⁸). The mononuclear cells in human atherosclerotic lesion express MCP-1 (**Fig. 4**).

Other than GM-CSF, many other pro-inflammatory cytokines such as TNF- α , interleukine-1 β , and IL-6 have been reported to enhance the MCP-1 expression in monocytes, endothelial cells, and fibroblasts³⁸). In addition, bacterial LPS stimulated monocytes/macrophages to produce MCP-1³⁹). Interestingly, LPS has also been reported to enhance Rho-me-





diated LFA-1 activation and AP-1-mediated MMP-1 production in monocytes⁴⁰⁻⁴¹). This MMP-1 gene transcription stimulated by LPS is not NF-kB (located at -20 to -10) dependent⁴⁰), and in our study mutation at the NF-kB-like site did not affect either basal activity or induction of the transcription by GM-CSF (unpublished data). There may thus exist a similar molecular mechanism in the signal transduction pathway of GM-CSF and LPS.

6. Expression of scavenger receptors and GM-CSF

A few reports describe GM-CSF regulation of scavenger receptor expression in monocytes. Treatment with GM-CSF

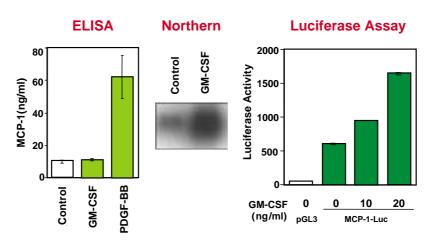


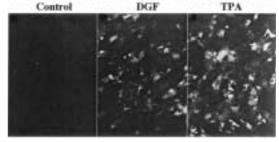
Figure 6 MCP-1 expression analysis by GM-CSF in monocytes.

down-regulates a number of binding sites for modified low density lipoprotein (LDL), and mRNA expression of type I and II scavenger receptor⁴²). The scavenger receptor CD163 in monocytes is down-regulated by treatment with GM-CSF and $IL-4^{43}-44$). In contrast, we found that inflammatory stimuli (histamine) stimulate monocytes to expression of lectin-like oxidused low-density lipoprotein receptor-1 (LOX-1)⁴⁵). We also found that transcriptional activity of Lox-1 is up-regulated by GM-CSF and IL-4, but not by PDGF-BB, in U937 monocytic cells (unpublished data). Since monocytic treatment with GM-CSF and IL-4 induces dendritic differentiation^{46~47}), GM-CSF may have a potential role of scavenger expression in monocytes and also in dendritic cells. Interestingly, monocyte-derived dendritic cells are activated by oxidized-LDL to increase HLA-DR, CD40 and CD86 expression⁴⁸).

7. GM-CSF effects on vascular smooth muscle cell

SMC are one of major constituents of vascular wall and participate in local inflammatory, immune responses⁴⁹ and MMP production⁵⁰. However, most studies have demonstrated PDGF effects on SMC and endothelial cells or fibroblasts in terms of MMP expression, migration and proliferation^{51~53} (**Fig. 7**). Based on our data, PDGF, but not GM-CSF, predominantly contributes to the functions of SMC to expression of MMP-12⁵⁴, while GM-CSF, but not PDGF, up-regulates monocytic MMP-12 expression¹⁷.

MMP-1 in cultured human smooth muscle cells Control DGF TPA



MMPs in human atherosclerotic lesions

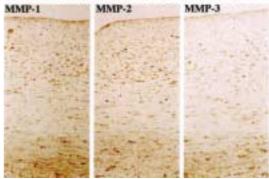


Figure 7 Expression of MMPs in SMC. Immunofluorescence staining:Synthesis of MMP-1 is stimulated in cultured human aortic smooth muscle cells by PDGF and TPA. Immunohistochemical staining: Expression of MMP-1, -2, and -3 is detected in human atherosclerotic intima.

One notable report is that GM-CSF stimulates SMC to express type VIII collagen⁵⁵, which is present in neonatal aorta rather than in adults' and is expressed at high levels in atherosclerotic lesion⁵⁶.

November, 25, 2003

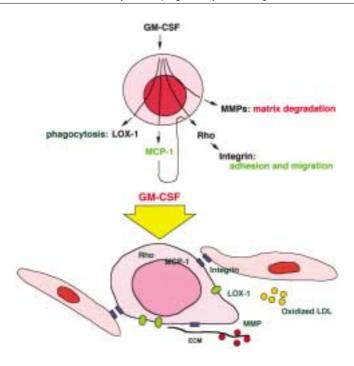


Figure 8 Multi-functional effects of GM-CSF in lesion environment of early stage of atherogenesis.

8. Source of GM-CSF in atherosclerotic lesion

In addition to the presense of GM-CSF in peripheral blood, monocytes/macrophages, endothelial, fibroblasts and smooth muscle cells can produce GM-CSF⁵⁷⁻⁵⁸). Monocyte and endothelial interaction, which is an initinal step of atherogenesis, stimulates both types of cells to express GM-CSF¹³). Modified-LDL induces GM-SCF expression in endothelial cells⁵⁹). Cultured human coronary SMCs constitutively express GM-CSF⁶⁰), and SMCs from internal mammary artery and aorta increase GM-CSF expression by treatment with IL-1 and TNF- α^{49} . GM-CSF localization in human atherosclerotic lesion is shown. Based on our data, multi-functions of GM-CSF are summarized in **Fig. 8**.

9. GM-CSF effects on angiogenesis and vasculogenesis

GM-CSF influences the migration and proliferation of human endothelial cells⁵⁸). Recent reports demonstrate that

GM-CSF enhances the endothelial cells mobilization from bone marrow- or peripheral blood-derived progenitor cells⁶¹⁻⁶²). This event is closely related to angioenesis and vasculogenesis⁶³⁻⁶⁴). Especially in the situation of arteriogenesis (collateral development from pre-existing arterioles) circulating monocytes adhere to the endothelium and play an important role in remodeling of the arterioles⁶⁴). Furthermore, GM-CSF is a strong arteriogenic factor, which regulates monocytes proliferation and life span⁶⁴⁻⁶⁵). The monocytes are also the predominant source of growth factors and proteolytic enzymes like MMP that regulate migration of SMC⁶⁵). In fact, promotion of collateral growth by GM-CSF has been reported in human coronary artery disease⁶⁶).

References

- 1) Ross R (1999). Atherosclerosis-An inflammatory disease. New Engl J Med **340**: 115-126.
- Reape TJ, and Groot PHE (1999). Chemokines and atherosclerosis. Atherosclerosis 147: 213-225.

- Watanabe T, Haraoka S, and Shimokama T (1996). Inflammatory and immunological nature of atherosclerosis. Int J Cardiol 54: S51-60.
- Hansson GK (1997). Cell-mediated immunity in atherosclerosis. Curr Opin Lipidol 8: 301-311.
- Sasaguri T, Arima N, Tanimoto A, Shimajiri S, Hamada T, Sasaguri Y (1998). A role of interleukin 4 in production of matrix metalloprotainase 1 by human aortic smooth muscle cells. Atherosclerosis 139: 247-253.
- 6) Zhou X, Paulsson G, Stemme S, Hansson GK (1998). Hypercholesterolemia is associated with a T helper (Th)1/Th2 switch of the autoimmune response in atherosclerotic apo E-knockout mice. J Clin Invest 101: 1717-1725.
- Folcik VA, Aamir R, Cathcart MK (1997). Cytokine modulation of LDL oxidation by activated human monocytes. Arterioscler Thromb Vasc Biol 17: 1954-1961.
- Ross R (1993). The pathogenesis of atherosclerosis: a perspective of the 1990s. Nature 362: 801-808.
- Yamada Y, Doi T, Hamakubo T, and Kodama T (1998). Scavenger receptor family proteins: roles for atherosclerosis, host defence. Cell Mol Life Sci 54: 628-640.
- Butcher EC (1991). Leukocyte-endothelial cell recognition: Three (or more) steps to specificity and diversity. Cell 67: 1033-1036.
- Hamilton JA (2002). GM-CSF in inflammation and autoimmunity. Trends Immunol 23: 403-408.
- Gasson JC (1991). Molecular physiology of granulocytemacrophage colony-stimulating factor. Blood 77: 1131-1145.
- 13) Takahashi M, Kitagawa S, Masuyama J, Ikeda U, Kasashara T, Takahashi Y, Furukawa Y, Kano S, and Shimada K (1996). Human monocyte-endothelial cell interaction induces synthesis of granulocyte-macrophage colony-stimulating factor. Circulation **93**: 1185-1193.
- 14) Sugiyama S, Okada Y, Sukhova GK, Virmani R, Heinecke JW, and Libby P (2001). Macrophage myeloperoxidase regulation by granulocyte macrophage colony-stimulating factor in human atherosclerosis and implications in acute coronary syndromes. Am J Pathol 158: 879-891.
- 15) Biwa T, Sakai M, Shichiri M, and Horiuchi S (2000). Granulocyte/macrophage colony-stimulating factor plays an essential role in oxidized low density lipoprotein-induced macrophage proliferation. J Atheroscler Thromb 7: 14-20.
- 16) Wang J, Wang S, Lu Y, Weng Y, Gown AM (1994). GM-CSF and M-CSF expression is associated with macrophage proliferation in progeressing and regressing rabbit athero-

matous lesions. Exp Mol Pathol 61: 109-118.

- 17) Wu L, Tanimoto A, Murata Y, Fan J, Sasaguri Y, and Watanabe T (2001). Induction of human matrix metalloproteinase-12 gene transcriptional activity by GM-CSF requires the AP-1 binding site in human U937 monocytic cells. Biochem Biophys Res Commun 285: 300-307.
- 18) Shang XZ, and Issekutz AC (1999). Enhancement of monocyte transendothelial migration by granulocyte-macrophage colony-stimulating factor: requirement for chemoattractant and CD11a/CD18 mechanisms. Eur J Immunol 29: 3571-3582.
- 19) Hamilton JA, Whitty GA, Stanton H, Wojta J, Gallichio M, McGrath K, Ianches G (1993). Macrophage colonystimulating factor and granulocyte-macrophage colonystimulating factor stimulate the synthesis of plasminogenactivator inhibitors by human monocytes. Blood 82: 3616-3621.
- 20) Hart PH, Vitti GF, Burgess DR, Whity GA, Royston K, Hamilton JA (1991). Activation of human monocytes by granulocyte-macrophage colony-stimulating factor: increased urokinase-type plasminogen activator activity. Blood 77: 841-848.
- 21) Schmitz AAP, Govek EE, Bottner B, and Van Aelst L (2000). Rho GTPases: Signaling, migration, and invasion. Exp Cell Res 261: 1-12.
- 22) Bevilacqua MP, and Nelson RM (1993). Selectins. J Clin Invest 91: 379-387.
- 23) Wojciak-Stothard B, Williams L, and Ridley AJ (1999). Monocyte adhesion and spreading on human endothelial cells is dependent on Rho-regulated receptor clustering. J Cell Biol 145: 1293-1307.
- 24) Baggiolini M, Dewald B, and Moser B (1997). Human chemokines: An update. Ann Rev Immunol 15: 675-705.
- 25) Kwon EM, Raines MA, Blenis J, and Sakamoto KM (2000). Granulocyte-macrophage colony-stimulating factor stimulation results in phosphorylation of camp response elementsbinding protein through activation of pp90RSK. Blood 95: 2552-2558.
- 26) Luscinskas FW, Kansas GS, Ding H, Pizcueta P, Schleiffenbaum BE, Tedder TF, and Gimbrone Jr MA (1994). Monocyte rolling, arrest and spreading on IL-4 activated vascular endothelium under flow is mediated via sequential action of L-selectin, β1 integrins, and β2 integrins. J Cell Biol **125**: 1417-1427.
- 27) Weber C, and Springer TA (1998). Interaction of very late antigen-4 with VCAM-1 supports transendothelial chemo-

taxis of monocytes by facilitating lateral migration. J Immunol 161: 6825-6834.

- 28) Goetzl EJ, Banda MJ, and Leppert D (1996). Matrix metalloproteinases in immunity. J Immunol 156: 1-4.
- 29) Zhang Y, Mccluskey K, Fujii K, and Wahl LM (1998). Differential regulation of monocyte matrix metalloproteinase and TIMP-1 production by TNF-α, granulocyte-macrophage CSF, and IL-1β through prostaglandin-dependent and -independent mechanisms. J Immunol 161: 3071-3076.
- 30) Matsumoto S, Kobayashi T, Katoh M, Saito S, Ikeda Y, Kobori M, Masuho Y, and Watanabe T (1998). Expression and localization of matrix metalloproteinase-12 in the aorta of cholesterol-fed rabbits. Relationship to lesion development. Am J Pathol 153: 109-119.
- 31) Sasaguri Y, Tanimoto A (2002). Extracellular matrix and matrix metalloproteinases in atherosclerosis. Connective Tissue 34: 327-336.
- 32) Janowska-Wieczorek A, Marquez LA, Nabholtz JM, Cabuhat ML, Montano J, Chang H, Rozmus J, Russell JA, Edwards DR, and Turner AR (1999). Growth factors and cytokines upregulate gelatinase expression in bone marrow CD34⁺ cells and their transmigration through reconstituted basement membrane. Blood **93**: 3379-3390.
- 33) Benbow U, and Brinckerhoff CE (1997). The AP-1 site and MMP gene regulation: What is all the fuss about? Martix Biol 15: 519-526.
- 34) Major TC, Liang L, Lu X, Rosebury W, Bocan TM (2002). Extracellular matrix metalloproteinase inducer (EMMPRIN) is induced upon monocyte differentiation and is expressed in human atheroma. Arterioscler Thromb Vasc Biol 22: 1200-1207.
- 35) Funayama H, Ikeda U, Takahashi M, Sakata Y, Kitagawa S, Takahashi Y, Masuyama J, Furukawa Y, Miura Y, Kano S, Matsuda M, and Shimada K (1998). Human monocyte-endothelial cell interaction induces platelet-derived growth factor expression. Cardiovasc Res 37: 216-224.
- 36) Takahashi M, Masuyama J, Ikeda U, Kasahara T, Kitagawa S, Takahashi Y, Shimada K, and Kano S (1995). Induction of monocyte chemoattractant protein-1 synthesis in human monocyte during transendothelial migration in vitro. Circ Res 76: 750-757.
- 37) Selgas R, Fernandez de Castro M, Jimenez C, Carcamo C, Contreras T, Bajo MA, Vara F, and Corbi A (1996). Immunomodulation of peritoneal macrophages by granulocyte-macrophage colony-stimulating factor in humans. Kidney Int 50: 2070-2078.

- 38) Biswas P, Delfanti F, Bernasconi S, Mengozzi M, Cota M, Polentarutti N, Mantovani A, Lazzarin A, Sozzani S, and Poli G (1998). Interleukin-6 induces monocyte chemotactic protein-1 in peripheral blood mononuclear cells and in the U937 cell line. Bood 91: 258-265.
- 39) Kopydlowski KM, Salkowski CA, Cody MJ, Van Rooijen N, Major J, Hamilton TA, and Vogel SN (1999). Regulation of macrophage chemokine expression by lipopolysaccharide in vitro an in vivo. J Immunol 163: 1537-1544.
- 40) Pierce RA, Sandefur S, Doyle GA, Welgus HG. Monocytic cell type-specific transcriptional induction of collagenase. J Clin Invest 1996; 97: 1890-1899
- Hmama Z, Knutson KL, Herrera-Velit P, Nandan D, and Reiner NE (1999). Monocyte adherence induced by lipopolysaccharide involves CD14, LFA-1, and cytohesin-1. J Biol Chem 274: 1050-1057.
- 42) Van der Kooij MA, Morand OH, Kempen HJ, Van Berkel TJ (1996). Decrease in scavenger receptor expression in human monocyte-derived macrophages treated with granulocyte macrophage colony-stimulating factor. Arterioscler Thromb Vasc Biol 16: 106-114.
- 43) Ritter M, Buechler C, Langmann T, Orso E, Klucken J, Schmitz G (1999). The scavenger receptor CD163: regulation, promoter structure and genomic organization. Pathobiology 67: 257-261.
- 44) Buechler C, Ritter M, Orso E, Langmann T, Kluken J, Schmitz G (2000). Regulation of scavenger receptor CD163 expression in human monocytes and macrphages by proand anti-inflammatory stimuli. J Leukoc Biol 67: 97-103.
- 45) Tanimoto A, Murata Y, Nomaguchi M, Kimura S, Arima N, Xu H, Hamada T, Sasaguri Y (2001). FEBS letters 508: 345-349.
- 46) Herbst B, Kohler G, Mackensen A, Veelken H, Lindemann A (1998). GM-CSF promotes differentiation of a precursor cell of monocytes and Langerhans-type dendritic cells from CD34⁺ haematopoietic progenitor cells. Br J Haematol 101: 231-241.
- 47) Meierhoff G, Krause SW, Andreesen R (1998). Comparative analysis of dendritic cells derived from blood monocytes or CD34⁺ hematopoietic progenitor cells. Immunobiology **198**: 501-513.
- 48) Alderman CJ, Bunyard PR, Chain BM, Foreman JC, Leake DS, Katz DR (2002). Effects of oxidized low density lipoprotein on dendritic cells: a possible immunoregulatory component of the atherogenic micro-environment? Cardiovasc Res 55: 806-819.

- 49) Filonzi EL, Zoellner H, Stanton H, Hamilton JA (1993). Cytokine regulation of granulocyte-macrophage colony stimulating factor and macrophage colony-stimulating factor production in human arterial smooth muscle cells. Atherosclerosis 99: 241-252.
- 50) Sasaguri Y, Murahashi N, Sugama K, Kato S, Hiraoka K, Satoh T, Isomoto H, Morimatsu M (1994). Developmentrelated changes in matrix metalloproteinase expression in human aortic smooth muscle cells. Lab Invest 71: 261-269.
- 51) Stringa E, Knauper V, Murphy G, and Gavrilovic J (2000). Collagen degradation and platelet-derived growth factor stimulate the migration of vascular smooth muscle cells. J Cell Science 113: 2055-2064.
- 52) Yanagi H, Sasaguri Y, Sugama K, Morimatsu M, and Nagase H (1992). Production of tissue collagenase (matrix metalloproteinase 1) by human aortic smooth muscle cells in response to platelet-derived growth factor. Atherosclerosis 91: 207-216.
- 53) Sasaguri Y, Yanagi H, Nagase H, Nakano R, Fukuda S, Morimatsu M (1991). Collagenase production by immortalized human aortic endothelial cells infected with simian virus 40. Virchows Archive B Cell Pathol 60: 91-95.
- 54) Wu L, Tanimoto A, Murata Y, Sasaguri T, Fan J, sasaguri Y, Watanabe T (2003). Matrix metalloproteinase-12 gene expression in human vascular smooth muscle cells. Genes Cells 8: 225-234.
- 55) Plenz G, Reichenberg S, Koenig C, Rauterberg J, Deng MC, Baba HA, Robenek H (1999). Granulocyte-macrophage colony-stimulating factor (GM-CSF) modulates the expression of type VIII collagen mRNA in vascular smooth muscle cells and both are codistributed during atherogenesis. Arterioscler Thromb Vasc Biol **19**: 1658-1668.
- 56) MacBeath JRE, Kielty CM, Shuttleworth CA (1996). Type VIII collagen is a product of vascular smooth-muscle cells in development and disease. Biochem J 319: 993-998.
- 57) Sallerfors B (1994). Endogenous production and peripheral blood levels of granulocyte macrophage (GM-) and granulocyte (G-) colony-stimulating factors. Leuk Lymphoma 13: 235-247.
- 58) Bussolino F, Wang JM, Defilippi P, Turrini F, Sanavio F, Edgell CJ, Aglietta M, Arese P, Mantovani A (1989). Granu-

locyte- and granulocyte-macrophage-colony stimulating factors induce human endothelila cells to migrate and proliferate. Nature **337**: 471-473.

- 59) Rajavashisth TB, Andalibi A, Territo MC, Berliner JA, Navab M, Fogelman AM, Lusis AJ (1990). Induction of endothelial cell expression of granulocyte and macrophage colony-stimulating factors by modified low-density lipoproteins. Nature **344**: 254-257.
- 60) Plenz G, Koenig C, Severs NJ, Robenek H (1997). Smooth muscle cells express qranulocyte-macrophage colony-stimulating factor in the undiseased and athensclerotic human coronary artery. Arterioscler Thromb Vasc Biol 17: 2489-2499.
- 61) Fischmeister G, Gadner H (2000). Granulocyte colonystimulating factor versus granulocyte-macrophage colony stimulating factor for collection of peripheral blood progenitor cells from healthy donors. Curr Opin Hematol 7: 150-155.
- 62) Takahashi T, Kalka V, Masuda H, Chen D, Silver M, Kearney M, Magner M, Isner JM, Asahara T (1999). Ischemia- and cytokine-induced mobilization of bone marrow-derived endothelial progenitor cells for neovascularization. Nat Med 5: 434-438.
- 63) Kalka C, Asahara T, Krone W, Isner JM (2000). Angionenesis and vasculogenesis. Therapeutic strategies for stimulation of postnatal neovascularization. Herz 25: 611-622.
- 64) Buschmann IR, Hoefer IE, van Royen N, Katzer E, Braun-Dulleaus R, Heil M, Kostin S, Bode C, Schaper W (2001). GM-CSF: a strong arteriogenic factor acting by amplification of monocyte function. Atherosclerosis 159: 343-356.
- 65) Scholtz D, Cai WJ, Schaper W (2001). Arteriogenesis, a new concept of vascular adaptation in occlusive disease. Angiogenesis 4: 247-257.
- 66) Seiler C, Pohl T, Wustmann K, Hutter D, Nicolet PA, Windecker S, Eberli FR, Meier B (2001). Promotion of collateral growth by granulocyte-macrophage colony stimulating factor in patients with coronary artery disease: a randomized, double-blind, placebo-controlled study. Circulation 104: 2012-2017.